

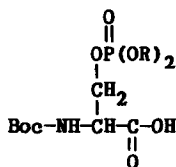
**SOLID-PHASE SYNTHESIS OF AN O-PHOSPHOSERYL-CONTAINING PEPTIDE  
 USING PHENYL PHOSPHOROTRIESTER PROTECTION**

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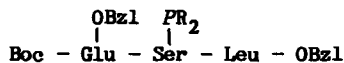
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**ABSTRACT:** The solid-phase synthesis of  $TFA \cdot H_2-Glu-PSer-Leu$  **1** is reported via incorporation of the protected phosphoroamino acid  $N_\alpha$ -tert-butoxycarbonyl-O-(diphenylphosphoro)-L-serine **2b**.

In our earlier studies <sup>1,2</sup>, we reported the solution-phase synthesis of Glu-PSer-Leu via a novel strategy which specifically incorporated a protected O-phosphoserine derivative into a peptide sequence, thereby, uniquely determining the position of the O-phosphoserine residue. This strategy involved the initial synthesis of the protected tripeptides **6a** and **6b** using **2a**<sup>3</sup> and **2b**<sup>2,4</sup> respectively followed by the hydrogenolytic removal of the benzyl or phenyl phosphorotriester protecting groups.



**2**    a) R = Bzl  
       b) R = Ph



**6**    a) R = Bzl  
       b) R = Ph

We now report the use of **2b**<sup>5</sup> in the synthesis of  $TFA \cdot H_2-Glu-PSer-Leu$  **1** via Merrifield solid-phase methodology<sup>6</sup>. By successive DCCI-HOBt couplings<sup>6</sup> of Boc-Ser[PPh<sub>2</sub>]-OH **2b** and Boc-Glu[OBzl]-OH **4** with 1.2g of Boc-Leu-Resin **3**<sup>7</sup> (1.0 mmol/g) and 40% TFA/CH<sub>2</sub>Cl<sub>2</sub> for cleavage of the Boc group from intermediate resin-bound peptides, 1.7g of resin-bound peptide **5** was obtained.

The  $^{13}\text{C}$  nmr spectrum of 5 in  $\text{CDCl}_3$ <sup>8</sup> (shown in Fig. 1) was found to be markedly similar to the  $^{13}\text{C}$  nmr spectrum obtained for Boc-Glu[OBzl]-Ser[PPh<sub>2</sub>]-Leu-OBzl 6b<sup>2</sup> (Fig. 2) and displayed the characteristic phosphorus-coupled doublet signals for the  $\alpha$ - and  $\beta$ -carbons of the O-(diphenylphosphoro)seryl residue at 53.3 and 67.8 ppm (each  $J_{\text{PC}}$  5.9 Hz). The  $^{31}\text{P}$  nmr spectrum of 5 contained a single broad resonance at -11.7 ppm<sup>9</sup> and compared favourably with a value of -12.1 ppm<sup>9</sup> obtained for 6b<sup>2</sup>.

Hydrogenolysis of 5 (1.20g) in 40% TFA/DMF<sup>10</sup> with  $\text{Pd}(\text{OAc})_2$ <sup>11</sup> simultaneously cleaved the peptide from the resin and removed the glutamyl benzyl and N-terminal Boc protecting groups<sup>12</sup>. A final hydrogenolytic treatment of the crude O-(diphenylphosphoro)tripeptide 7 in 40% TFA/AcOH with  $\text{PtO}_2$  (1.0 eq/phenyl group) cleaved the phenyl phosphotriester groups and gave 1 (250 mg) as a white solid after trituration with diethyl ether. Chromatographic purification of 1<sup>13</sup> was not required since  $\text{C}_{18}$  RP-HPLC analysis ( $\mu$  Bondapak C18, 0-50%  $\text{CH}_3\text{CN}/0.1\%$  TEAP pH 4.0, 1 ml/min, 214 nm) showed it to be homogeneous (> 95% pure)<sup>14</sup>. In addition to amino acid analysis of the acid hydrolysate of 1 giving the constituent amino acid residues in the expected molar ratios (Glu<sub>1</sub> 1.00, Ser<sub>1</sub> 0.95, Leu<sub>1</sub> 0.95), chiral evaluation of these amino acid residues via  $\text{C}_{18}$  RP-HPLC analysis of their L-Leucyl dipeptides established them to be racemization free.

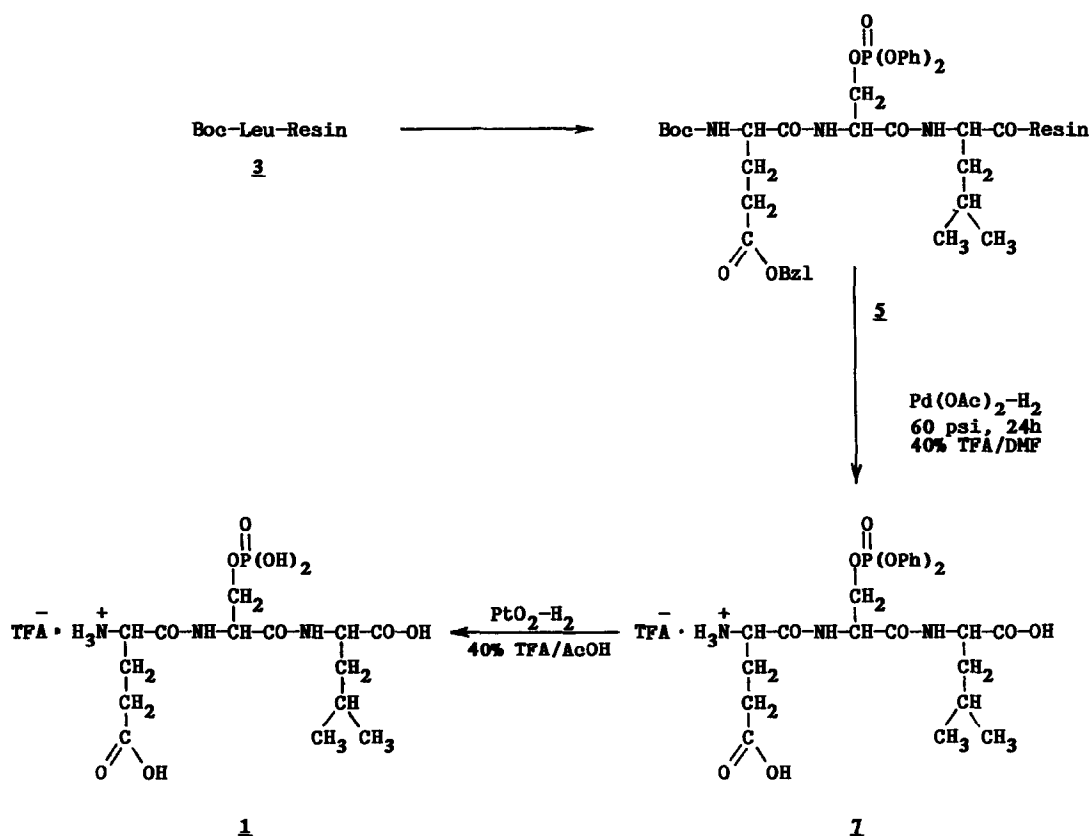


Fig. 1

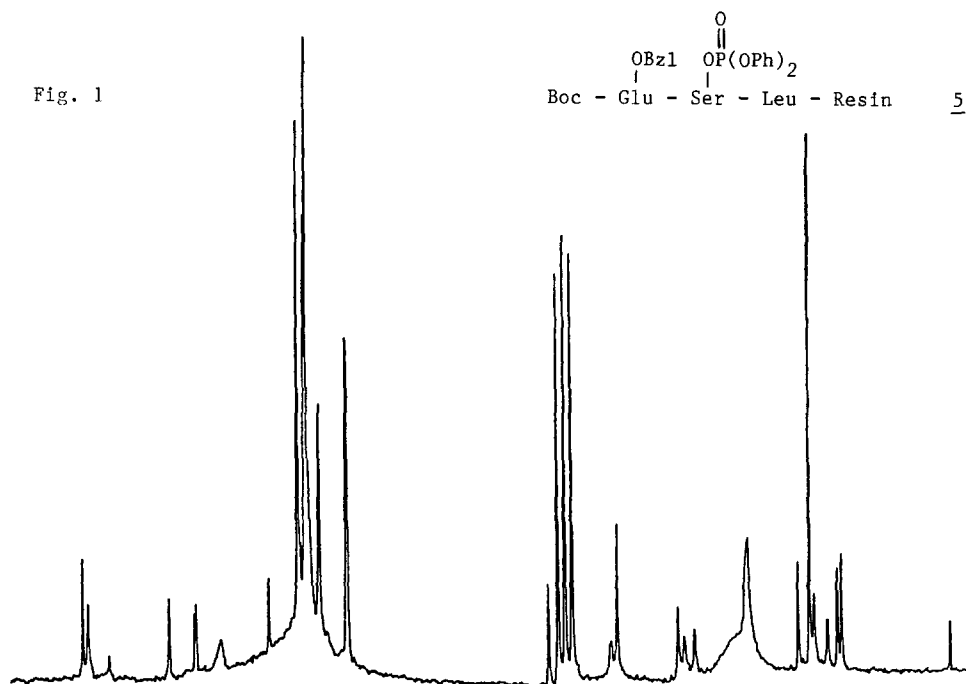
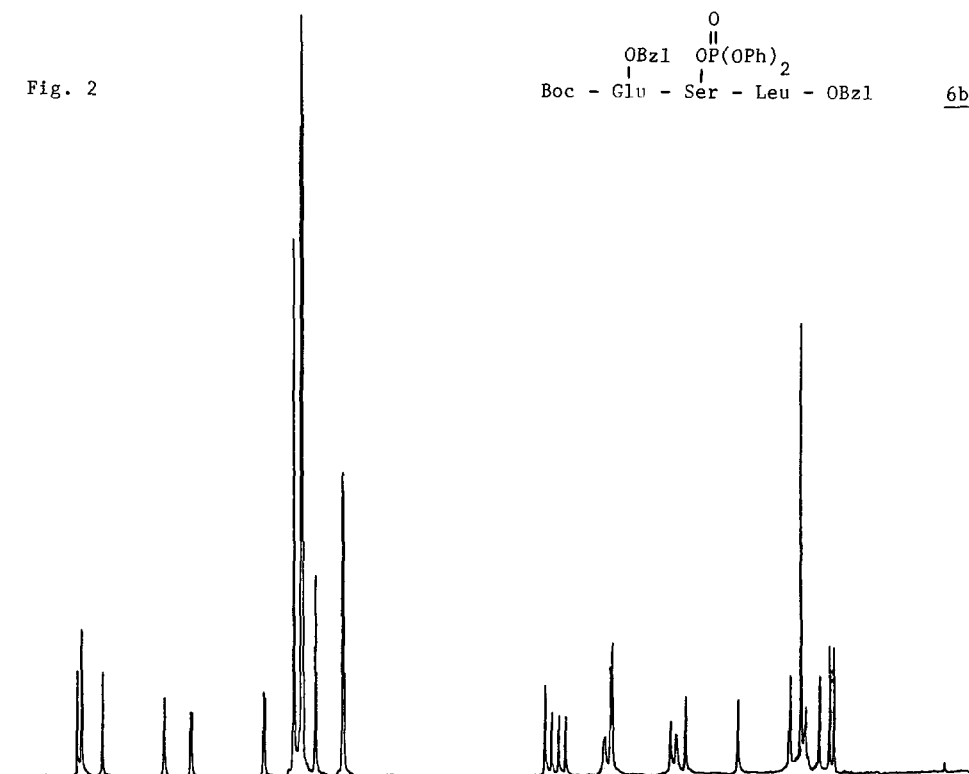


Fig. 2



The efficient synthesis of TFA · H<sub>2</sub>-Glu-PSer-Leu 1 using Boc-Ser[Ph<sub>2</sub>]-OH 2b demonstrates that N-protected O-(disubstitutedphosphoro)serine derivatives are readily adapted to solid-phase peptide synthesis and indicates that the synthesis of longer and more complex O-phosphoserine-containing peptides to be a practical proposition.

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**NOTES AND REFERENCES:**

1. P.F. Alewood, J.W. Perich and R.B. Johns, *Tetrahedron Lett.*, **25**, 987, (1984).
2. J.W. Perich, P.F. Alewood and R.B. Johns, *Tetrahedron Lett.*, preceding publication.
3. P.F. Alewood, J.W. Perich and R.B. Johns, *Aust. J. Chem.*, **37**, 429, (1984).
4. J.W. Perich, P.F. Alewood and R.B. Johns, *Synthesis*, accepted for publication (1985).
5. 2a could not be satisfactorily applied to solid-phase synthesis due to the acidolytic sensitivity of benzyl phosphotriester groups to 40% TFA/CH<sub>2</sub>Cl<sub>2</sub>.
6. R.S. Hodges and R.B. Merrifield, *Anal. Biochem.*, **65**, 241, (1975).
7. Prepared by the esterification of Boc-Leu-O<sup>-</sup> <sup>+</sup>Cs with chloromethylated styrene-1% divinylbenzene resin, 1.04 mmol Cl/g resin (FLUKA).
8. <sup>13</sup>C nmr δ (CDCl<sub>3</sub>) 5: 21.9, 22.7, 24.7, 27.4, 28.3, 30.6, 40.6, 51.2, 53.3 (d, J<sub>PC</sub> = 5.9 Hz), 57.5, 66.6, 67.8 (d, J = 5.9 Hz), 80.34, 120.1 (d, J<sub>PC</sub> = 4.9 Hz), 125.6, 128.2, 128.6, 129.9, 135.8, 150.3 (d, J<sub>PC</sub> = 7.3 Hz), 167.7, 171.9 and 173.0 ppm.
9. Relative to ext. 85% H<sub>3</sub>PO<sub>4</sub>.
10. Hydrogenolysis of 5 in neat DMF gave rise to an unknown side-product.
11. J.M. Schlatter, R.H. Mazur and O. Goodmanson, *Tetrahedron Lett.*, **23**, 2851, (1977).
12. Under these conditions, <sup>31</sup>P nmr spectroscopy showed ~5% cleavage of a phenyl group occurred.
13. Identical in all respects with TFA · Glu-PSer-Leu prepared via solution-phase peptide synthesis<sup>2</sup>.
14. C<sub>18</sub> RP-HPLC analysis of 1 confirmed quantitative incorporation of 2b had occurred during peptide synthesis since it was not contaminated with either TFA · PSer-Leu or leucine.

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